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1 **Optimization of Conditions for the Cleaning of overpaint from the Stone Carvings of**  
2 **the Potala Palace, Tibet using enzyme-based cleaning agents**

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20 **ABSTRACT**

21 This work focuses on the study of the removal of overpaint from the stone carvings  
22 of the Potala Palace, Tibet. The removal of historical mortar and overpaint is a significant  
23 challenge for the conservation of stone carvings and requires careful optimization. Paints  
24 applied in the past were made of a mixture of hydraulic lime, water and milk, and are  
25 causing damage to the carvings of the Potala Palace. The protein content of lime-based  
26 paint samples collected from the Potala Palace was determined by enzyme-linked  
27 immunosorbent assay (ELISA). In order to optimize the removal of the overpaint without  
28 causing abrasion or other damage to the underlying stone carvings, we prepared a simulated  
29 paint with the same protein content. Removal rate and color difference were used to select  
30 the best cleaning agent from ionic surfactant, ethanol/aqueous solutions, five proteases and  
31 Twin-20 (non-ionic surfactant). With color difference and gloss as evaluation indexes, the  
32 influencing factors (temperature, pH, enzyme concentration, and enzyme hydrolysis time)  
33 were measured. Results suggest that alkaline protease was the most suitable cleaning agent  
34 for removing surface paint, and the best working condition is an enzymatic dosage of 1.92  
35 mg/mL under 42°C, pH of 9.5, with a hydrolysis time of 5.4 min. This method generated  
36 the desired color difference of 2.25 and the minimal gloss ( $0.1 \pm 0.1$  GU) on the underlying  
37 stone, with no significant variation from the predicted value ( $P < 0.05$ ). Before and after  
38 cleaning, the change in surface roughness was less than 10%. This study provided a  
39 feasibility plan for removing the paints from the outer wall of the Potala Palace by using  
40 enzymatic hydrolysis.

41 **Key words:** whitewash removal, the Potala Palace, Protein, Alkaline protease cleaning,  
42 Response surface methodology

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43 **1. Introduction**

44       The Potala Palace is Tibet's largest best-preserved palace-style fortress complex<sup>[1]</sup>.  
45       According to the local religious tradition, the walls of the Potala Palace are repainted  
46       with a lime-based paint every year. Locals typically combine milk, honey, and other  
47       materials with lime to manufacture paint, which is applied over the Potala Palace's  
48       outside walls to increase the consistency and stickiness of the paint<sup>[2]</sup>. Due to this  
49       tradition, layers of lime-based paint now covers the surface of originally unpainted stone  
50       carvings, which seriously affects the overall appearance of the Potala Palace<sup>[3]</sup>. The paint  
51       compromises both the legibility and the preservation of stone artifacts as paint obscures  
52       the surface of stones, and can create a barrier to moisture transport. Moreover, the paint  
53       covers up the cracks that easily expose the structural problems of the stone, resulting in  
54       missing the best restoration period. Therefore, an adequate cleaning strategy to remove  
55       the overpaint is necessary.

56       Lime-based mortars are primarily used as for masonry, jointing and plastering, while  
57       lime-based paints can be applied to stone as whitewash. Whitewash primarily consists of  
58       cementing materials (lime), water, and additives<sup>[4,5]</sup>. Several organic materials, such as  
59       milk<sup>[11-13]</sup>, animal glue<sup>[14]</sup>, egg white<sup>[15]</sup>, and blood<sup>[16,17]</sup> are often added to lime in  
60       whitewash to improve the mechanical strength and adhesion of the paint.

61       Physical removal of overpaint from stone may be accomplished with dry cleaning  
62       (laser cleaning and particle injection) and wet cleaning (high-pressure water cleaning and  
63       steam cleaning)<sup>[18]</sup>. Despite the extensive applications of these cleaning technologies,  
64       obvious limitations still remain. For example, laser cleaning can remove micron-level  
65       adhesions on the stone carvings surface, but its effect is mostly confined to specific cases

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66 such as dirt accumulation, along with very low efficiency and high cost<sup>[19,20]</sup>. Abrasive  
67 blasting may cause surface damage. Dust contamination in the operating environment must  
68 be controlled<sup>[21,22]</sup>. High-pressure water spraying is challenging to control, resulting in  
69 damage to substrates that may result in discoloration, unevenness, flaking, cracks, etc<sup>[23]</sup>.  
70 The involvement of water will introduce moisture into the artifacts, resulting in the transfer  
71 of contaminants<sup>[24]</sup>. The high-temperature steam employed can cause uneven thermal  
72 expansion inside and outside the stone carvings artifacts and result in stress damage<sup>[25]</sup>.

73       Mechanochemical cleaning is based on mechanical cleaning with the addition of  
74 chemical reagents such as redox, organic solvents, gels and bioactive materials to clean the  
75 surface of the artifacts<sup>[26-28]</sup>, which can be applied with adsorbent materials<sup>[29-32]</sup>. Chemical  
76 agents may endanger cultural relics and may lead to the development of secondary reactant  
77 that may damage the substrate<sup>[33,34]</sup>. By contrast, biological cleaning technologies have  
78 been proposed as safer for artifacts, the environment, and operators<sup>[35,36]</sup>. Using  
79 microorganisms and enzymes to target molecules within overpaint is an effective  
80 alternative to traditional cleaning technologies. By using enzymolysis to target protein in  
81 paint<sup>[37,38]</sup>, Abdelaal et al. showed that protease can be safely used to remove unwanted  
82 substances in a controlled manner<sup>[39]</sup>. Proteases provide a powerful tool that enables many  
83 fragile artifacts to be cleaned and repaired efficiently and quickly<sup>[40]</sup>.

84       Our study focuses on the optimization of the remove the overpaint from the surface  
85 of stone carvings by targeting the organic binder used in the paint mixture. Considering  
86 that the main component of the paint used in the Potala Palace is a protein-based compound,  
87 we selected protease as a cleaning agent because it can specifically bind proteins in milk.  
88 Influencing factors (temperature, pH, enzyme concentration, and enzymatic hydrolysis

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89 time) were optimized with the Box-Behnken Design to identify the best cleaning conditions.  
90 To evaluate the cleaning result, we compared the surface roughness values of the blank  
91 stones with the stones cleaned under ideal conditions.

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## 93 **2. Research aims**

94 The Potala Palace has great significance, but is subject to the hazards of natural  
95 conditions and the past application of paint and mortar which greatly affects the  
96 preservation value of stone carvings. The aim of this study is to find a safe, efficient,  
97 inexpensive, and environmentally friendly cleaning method to remove overpaint from the  
98 surface of stone carvings, and to fill the gap in the cleaning technology of stone carvings.

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99 **3. Materials and methods**

100 **3.1 Materials and reagents**

101 Milk with a protein concentration of 0.03 g/mL was procured by Inner Mongolia  
102 Mengniu Dairy (Group) Co., Ltd.(China). Hydrated lime powder was purchased from  
103 Shishun Limestone Mine Co., Ltd., Mingshan District, Benxi City (China). Enzyme-  
104 linked immunosorbent assay kit was bought from Shanghai Fusheng Industrial Co.,  
105 Ltd.(China). Alkaline protease containing  $2.0 \times 10^5$  U/g was manufactured by  
106 Novozymes Biotechnology Co., Ltd.(China). Papain enzyme-containing  $3.0 \times 10^3$  U/g  
107 was manufactured by Sanland Chem International Co., Ltd.(China). The tryptic  
108 enzyme-containing  $1.0 \times 10^5$  U/g was made by Shanghai Aladdin Biochemical  
109 Technology Co., Ltd. (China). Bromelain containing  $1.7 \times 10^3$  U/g was obtained from  
110 Nanning Pangbo Bioengineering Co., Ltd.(China). Sodium hydroxide and  
111 hydrochloric acid (analytically pure) were purchased from Tianjin Benchmark  
112 Chemical Reagent Co., Ltd.(China). Sodium citrate, ethanol, and Tween 20  
113 (analytically pure) were supplied by Shanghai Eon Chemical Technology Co.,  
114 Ltd.(China).

115 **3.2 Experimental design**

116 **3.2.1 Paint samples**

117 The paint samples were taken from the surface of the stone sculptures at the Potala  
118 Palace. In Fig. 1, 5 g of samples were taken from each sampling point in the outer,  
119 middle, and bottom layers for the subsequent analysis. Fig. 2 shows the field images of  
120 the sampling points in H10-E39. (a: outer layer, b: middle layer, c: bottom layer). From  
121 Fig. 3, a total of 10 paint samples were acquired.

122 **3.2.2 Determination of protein content of the paint**

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123 The protein crude extract was obtained by first mixing 4 mg of samples with 0.1  
124 ml of encapsulation buffer and then extracted at 60°C for 10 min, followed by  
125 centrifugation at 3000 g for 10 min at 4°C. 50 µL of bovine casein was first placed into  
126 a 96-well ELISA plate, then 40 µL of sample dilution and 10 µL of the sample were  
127 added to the sample wells sequentially. The enzyme labeling plate was wrapped in  
128 sealing film and then warmed at 37°C for 30 min. The wash solution was then poured  
129 onto the enzyme labeling plate, which was then removed and left for 30 s before being  
130 repeated 5 times. Each well was spiked with 50 µL of enzyme standard reagent except  
131 for the blank. After repeated warming and washing operations, 50 µL each of  
132 chromogenic agents (A and B) were added to the enzyme labeling plate and left for 10  
133 min at 37°C and protected from light. Finally, the stopping solution was applied to the  
134 enzyme labeling plate<sup>[41]</sup>.

### 135 **3.2.3 Preparation of simulated stones**

136 According to the protein content of the samples measured in 3.3.2, the simulated  
137 paint was made of milk, water and lime powder, in which the milk was 5% of the liquid,  
138 and the solid ratio of liquid (milk + water) was 0.61<sup>[42]</sup>. The paint was stirred until  
139 homogeneous and then applied to the surface of the original stones of equal size (5 ×  
140 5 × 2 cm), which were provided by the Potala Palace Administration and were made of  
141 the same material as the stone artifacts. Considering the temperature and UV intensity  
142 of Lhasa, the simulated stones were aged in a UV weathering test box (Foshan Huahe  
143 Zhensen Testing Machine Manufacturing Co., Ltd.(China)) in order to achieve the  
144 effect of sunlight exposure for one year in a shorter period of time. 80 samples were  
145 prepared by applying a paint (with protein content 0.86%) on unpainted stones. These  
146 were then placed in the ultraviolet aging box, with a UV exposure of 0.68 w/cm<sup>2</sup> and  
147 an irradiation time of 11 days. These stones are known as simulated stones.

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### 148 3.2.4 Screening of cleaning agents

149 The optimal cleaning agent was selected from organic reagents (1 mol/L sodium  
150 citrate solution and 95% ethanol), distilled water, different types and concentrations of  
151 proteases (bromelain, neutral protease, papain, alkaline protease, and tryptic enzyme),  
152 and a non-ionic surfactant (Tween 20). To compare the effect of different cleaning  
153 agents on stones without paint, a simple corrosion test was carried out. Stones were  
154 soaked in cleaning agents 2 hours, and then washed in water and dried. The difference  
155 in mass was calculated by comparing the stone before and after soaking. <sup>[43]</sup>. The  
156 damage to the stone surface due to the corrosive action of the cleaning agent was  
157 calculated according to the Eq. (1):

$$158 W(\%)=(m_1-m_2)/m_1\times 100\%.(1)$$

159 Where W is the corrosion rate;  $m_1$  is the mass of the original stone; and  $m_2$  is the mass  
160 of the cleaned stone.

### 161 3.2.5 Single factor experiments

162 The role of alkaline protease in cleaning the paint from stones was investigated  
163 using single factor experiments with four independent variables (temperature, pH,  
164 enzyme concentration, and enzymatic hydrolysis time). The simulated stones were  
165 wetted with paper towels and cling film for different times (3 min, 5 min, 9 min, and  
166 15 min) at various temperatures (30°C, 35°C, 40°C, and 45°C), pH (7, 8, 9, and 10),  
167 and enzyme concentration (1.0 mg/mL, 1.5 mg/mL, 2.0 mg/mL, and 2.5 mg/mL) and a  
168 soft brush was used to remove the softened paint. After the simulated stones were dried,  
169 color difference and gloss were measured<sup>[29]</sup>.

### 170 3.2.6 Optimization of alkaline protease for cleaning

171 Based on the results of the single factor experiments, we utilized a four-factor,  
172 three-level response surface analysis method using the Box-Behnken design. Table 1



173 shows the effect of temperature (A), pH (B), enzyme concentration (C), and enzymatic  
 174 hydrolysis time (D) on the cleaning effect using color difference and gloss as  
 175 assessment indices<sup>[44]</sup>.

176 Table1 Experimental range and levels of independent variables

Independent variables	Coded levels		
	-1	0	1
A: Temperature(°C)	35	40	45
B: pH	8	9	10
C: Enzyme concentration(mg/mL)	1.0	1.5	2.0
D: Enzymatic hydrolysis time (min)	3	5	7

177 **3.2.7 Determination of the color difference**

178 A colorimeter (CM-700d, Konica Minolta Co., Ltd.(China)) was utilized to  
 179 measure the standard samples, which were the original sample stones without paint, and  
 180 the obtained values were recorded as  $L^*_1$ ,  $a^*_1$ , and  $b^*_1$ . Then the simulated stones under  
 181 different cleaning conditions were examined using the colorimeter, and the values  
 182 obtained were recorded as  $L^*_2$ ,  $a^*_2$ , and  $b^*_2$ . Each of simulated stone surface was  
 183 measured at three points<sup>[45]</sup>. The color difference was calculated according to the Eq.(2):

184 
$$\Delta E = \sqrt{(\Delta L^*)^2 + (\Delta a^*)^2 + (\Delta b^*)^2}.(2)$$

185 Where  $\Delta L^*=L^*_2-L^*_1$  (brightness factor difference);  $\Delta a^*=a^*_2-a^*_1$  (red-green Index  
 186 Difference);  $\Delta b^*=b^*_2-b^*_1$  (yellow-blue index difference). For this work, the cleaning  
 187 effect is considered excellent ( $\Delta E \leq 3$ ); the cleaning effect is general ( $3 < \Delta E \leq 5$ ); and  
 188 cleaning experiments are failed ( $\Delta E > 5$ )<sup>[29]</sup>.

189 **3.2.8 Determination of gloss**

190 The change in surface morphology of the simulated stones before and after  
 191 cleaning under different conditions was presented in terms of gloss. The glossiness

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192 instrument (3nh Co., Ltd.(China)) was used to measure the original samples, and the  
193 obtained values were noted as  $GS_1$  (GU). The simulated stones under different cleaning  
194 conditions were examined and the values obtained were noted as  $GS_2$  (GU)<sup>[46]</sup>. Gloss  
195 was calculated according to the Eq.(3):

$$196 \quad GS = GS_1 - GS_2. \quad (3)$$

197 Where cleaning effect is excellent when GS is very small.

### 198 **3.2.9 Determination of surface roughness**

199 The surface roughness was used to check whether the cleaning affected the stone  
200 surface<sup>[47]</sup>. As the control group of three original stones, nine measuring points were  
201 selected on the surface of each original stone in three rows and three columns. The  
202 values of the original stones measured by the TR 200 surface analyzer (Beijing Jitai  
203 Keyi Testing Equipment Co., Ltd.(China)) were averaged and then recorded as  $Ra_1$ . The  
204 blank stones were coated with protein-containing paint, and treated under optimal  
205 cleaning conditions. Nine measuring points were selected on the surface of each painted  
206 stone. The values measured for the painted stones were averaged and then noted as  $Ra_2$ .  
207 The surface roughness was calculated using the Eq. (4):

$$208 \quad \Delta Ra = (Ra_1 - Ra_2) \times 100\%. \quad (4)$$

209 In this work, cleaning is considered to be excellent ( $\Delta Ra \leq 10\%$ ); cleaning effect  
210 is general ( $10\% < \Delta R \leq 15\%$ ); and cleaning experiments are deemed to have failed  
211 ( $\Delta R > 15\%$ )<sup>[48]</sup>.

### 212 **3.3 In-situ experiment**

213 During cleaning of stone carvings, the first step was to remove surface dirt from  
214 the paint. This was followed by the application of an alkaline protease under the best  
215 conditions to wet the stone carvings. The paint was removed using a soft brush dipped  
216 in clean water.

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217 **3.4 Statistical analyses**

218 The data obtained from the experiment were described by the mean and standard  
219 deviation. The experimental data were analyzed using Excel 2019 and IBM SPSS  
220 Statistics 25 software. Box-Behnken Design and variance analysis were completed by  
221 Design-Expert. V8.0.6 software. All experiments were performed three times in parallel.

222 **4. Results and discussion**

223 **4.1 Results and analysis of protein content in samples**

224 **4.1.1 Standard curve of casein**

225 We calculated casein content in the paint samples by ELISA. The standard curve  
226 is plotted in Fig. 4. The standard curve's determination coefficient ( $R^2$ ) was 0.9965,  
227 implying that the model was well fitted. The linear equation of the standard curve for  
228 the determination using ELISA was as follows:

229  $Y=0.0142X+0.0097$ . (5)

230 Where Y is the absorbance value of casein, and X is the concentration of casein solution.

231 **4.1.2 Protein content in paint samples**

232 The results of protein identification of 10 paint samples determined by ELISA are  
233 presented in Table 2, where some of the samples showed significant differences in  
234 protein content ( $P < 0.05$ ). After calculation, the protein percentage was estimated as  
235 0.68%.

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242 Table 2 Protein content in paint samples

Run	Name of samples	Protein concentration ( $\mu\text{g/mL}$ )
1	H10-E37 stone carving outer paint layer	206.75 $\pm$ 5.12 <sup>b</sup>
2	H10-E39 stone carving middle layer	181.85 $\pm$ 3.82 <sup>de</sup>
3	H10-E11 stone carving outside the double paint layer	183.20 $\pm$ 1.39 <sup>de</sup>
4	West side lower road outer paint layer	271.20 $\pm$ 4.69 <sup>a</sup>
5	H10-E11 stone carving bottom paint layer	177.09 $\pm$ 5.51 <sup>e</sup>
6	H10-E11 stone carving middle paint layer	178.95 $\pm$ 4.40 <sup>e</sup>
7	H10-E39 stone carving bottom paint layer	179.49 $\pm$ 3.61 <sup>e</sup>
8	H10-E39 stone carving outer paint layer	195.13 $\pm$ 3.51 <sup>c</sup>
9	H10-E37 stone carving bottom paint layer	188.62 $\pm$ 2.49 <sup>cd</sup>
10	H10-E11 stone carving outer paint layer	193.55 $\pm$ 4.49 <sup>c</sup>

243 **Notes:** Different letters within a column indicate significant differences at  $P < 0.05$

#### 244 4.2 Analysis of cleaning agents

245 According to Fig. 5, we found that sodium citrate solution (1 mol/L) and ethanol  
 246 (95%) were more corrosive to the surface of simulated stones<sup>[25]</sup>, while distilled water,  
 247 alkaline protease (1 mg/mL), and Tween 20 were less corrosive. In Fig. 6, although the  
 248 cleaning effects of distilled water and Tween 20 (0.3%) were not as great as alkaline  
 249 protease, Tween 20 (0.3%) could achieve the purpose of cleaning the paint from  
 250 simulated stones<sup>[49]</sup>. Therefore, we studied the cleaning effect by the combination of  
 251 alkaline protease (1 mg/mL) with Tween 20 (0.3%) as compared to alkaline protease (1  
 252 mg/mL) alone. The experimental results showed no significant difference ( $P < 0.05$ ) in  
 253 the cleaning effect of the two cleaning agents. In order to reduce the cost and achieve a

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254 better cleaning effect, alkaline protease was selected as the cleaning agent for simulated  
255 stones in this study.

### 256 **4.3 Optimization of the cleaning conditions with single-factor experiments**

#### 257 **4.3.1 Temperature**

258 We evaluated the color difference and gloss of painted stones at different  
259 temperatures ranging from 30°C to 45°C to observe how temperature affected the  
260 cleaning result. The results in Fig. 7a indicated that from 30°C to 40°C, both the color  
261 difference and gloss decreased sharply with increasing temperature. This phenomenon  
262 was due to the activation of the active gene of alkaline protease at lower temperature,  
263 resulting in the most efficient hydrolysis of casein in milk. The degree of casein  
264 hydrolysis reached a maximum value at the ideal temperature<sup>[50,51]</sup>. Above the ideal  
265 temperature, the removal of paint is compromised due to enzyme inactivation. Based  
266 on the cleaning effect, we set the optimal cleaning temperature at 40°C for subsequent  
267 tests.

#### 268 **4.3.2 pH**

269 We tested the color difference and gloss in the pH range of 7 to 10 to find the best  
270 pH for cleaning paint from stones with alkaline protease. [The results in Fig. 7b showed](#)  
271 [that when pH was 7-9, the color difference and gloss had a substantial negative](#)  
272 [association, but showed no significant changes when pH was further increased to 10.](#)  
273 The negative correlation could be explained by the most rapid hydrolysis rate of  
274 alkaline protease when working under the optimum pH range, which led to changes in  
275 color and gloss<sup>[52,53]</sup>. When exceeding the optimum pH range, changes in color and  
276 gloss were reduced due to the loss of enzyme activity. Considering the cleaning effect,  
277 we selected pH (9) as the most suitable starting point for subsequent tests.

#### 278 **4.3.3 Enzyme concentration**

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279 We investigated the color difference and gloss of painted stones with enzyme  
280 concentrations ranging from 1.0 mg/mL to 2.5 mg/mL to acquire adequate parameters  
281 to understand the influence of enzyme concentration on cleaning performance. Fig. 7c  
282 illustrates that the color difference and gloss gradually decreased through increasing the  
283 enzyme concentration. However, increasing enzyme concentration beyond 1.5 mg/mL  
284 eliminated the negative correlation and showed a smooth trend. The explanation was  
285 that as the enzyme concentration increased so did the hydrolysis of the paint binder.  
286 Later, when the enzyme concentration surpassed 1.5 mg/mL, a part of the alkaline  
287 protease could no longer bind to the proteins due to the saturation of most substrates<sup>[54]</sup>.  
288 According to the cleaning performance, we chose 1.5 mg/mL as the starting point for  
289 subsequent tests.

#### 290 **4.3.4 Application time**

291 We conducted experiments ranging from 3 min to 15 min to test how enzymatic  
292 hydrolysis time impacted cleaning efficiency. In Fig. 7d, both the color difference and  
293 gloss decreased steadily with increased enzymatic hydrolysis time from 3 min to 5 min.  
294 The two indexes gradually stabilized when the enzymatic hydrolysis time  
295 surpassed 5 min. This phenomenon could be ascribed to the fact that the catalytic  
296 site of alkaline protease decreased as the hydrolysis reaction proceeded<sup>[55]</sup>,  
297 which led to a declined protein hydrolysis level in the paint that generally  
298 maintained the color difference and the gloss of the simulated stones. Given  
299 the cleaning effect, we selected 5 min as the starting point for subsequent tests.

#### 300 **4.4 Optimization of the cleaning conditions**

301 After carrying out single-factor experiments, we used four factors with three levels  
302 and 29 different runs to compare the results of the combined treatments and the data  
303 obtained are recorded in Table 3.

Table 3 Box-Behnken response surface design and experimental results

Run	Factors				Responses	
	A(°C)	B	C(mg/mL)	D(min)	$\Delta E$	GS (GU)
1	-1(35)	-1(8)	0(1.5)	0(5)	7.15	1.1
2	1(45)	-1(8)	0(1.5)	0(5)	4.59	0.6
3	-1(35)	1(10)	0(1.5)	0(5)	6.29	1.0
4	145()	1(10)	0(1.5)	0(5)	3.07	0.3
5	0(40)	0(9)	-1(1.0)	-1(3)	5.66	0.8
6	0(40)	0(9)	1(2.0)	-1(3)	4.39	0.6
7	0(40)	0(9)	-1(1.0)	1(7)	4.67	0.6
8	0(40)	0(9)	1(2.0)	1(7)	2.86	0.2
9	-1(35)	0(9)	0(1.5)	-1(3)	6.60	1.0
10	1(45)	0(9)	0(1.5)	-1(3)	5.66	0.8
11	-1(35)	0(9)	0(1.5)	1(7)	5.38	0.8
12	1(45)	0(9)	0(1.5)	1(7)	3.07	0.3
13	0(40)	-1(8)	-1(1.0)	0(5)	6.38	0.9
14	0(40)	1(10)	-1(1.0)	0(5)	5.38	0.8
15	0(40)	-1(8)	1(2.0)	0(5)	4.67	0.6
16	0(40)	1(10)	1(2.0)	0(5)	2.75	0.4
17	-1(35)	0(9)	-1(1.0)	0(5)	6.21	0.9
18	1(45)	0(9)	-1(1.0)	0(5)	5.22	0.7
19	-1(35)	0(9)	1(2.0)	0(5)	5.66	0.8
20	1(45)	0(9)	1(2.0)	0(5)	4.06	0.5
21	0(40)	-1(8)	0(1.5)	-1(3)	6.05	0.9
22	0(40)	1(10)	0(1.5)	-1(3)	5.22	0.7
23	0(40)	-1(8)	0(1.5)	1(7)	4.03	0.5
24	0(40)	1(10)	0(1.5)	1(7)	2.35	0.2
25	0(40)	0(9)	0(1.5)	0(5)	2.90	0.3
26	0(40)	0(9)	0(1.5)	0(5)	2.90	0.2
27	0(40)	0(9)	0(1.5)	0(5)	3.23	0.2
28	0(40)	0(9)	0(1.5)	0(5)	2.90	0.3
29	0(40)	0(9)	0(1.5)	0(5)	2.56	0.4

305 Notes: A temperature; B pH; C enzyme dosage; D enzymatic hydrolysis time.

#### 306 4.4.1 Model fitting and statistical analysis

307 The results of the four factors and three levels are shown in Table 3. The results in

308 Table 4 and regression equations were obtained by multiple linear regression analysis

309 and quadratic multinomial fitting of the data in Table 3 with Design-Expert. V8.0.6  
310 software.

311  $Y_{\Delta E}=2.90-0.97A-0.65B-0.76C-0.94D-0.17AB-0.15AC-0.34AD-0.23BC-0.21BD-$   
312  $0.13CD+1.53A^2+0.90B^2+0.90C^2+0.65D^2$ . (6)

313  $Y_{GS}=0.28-0.20A-0.100B-0.13C-0.18D-0.050AB-0.025AC-0.075AD-0.025BC-$   
314  $0.025BD-0.050CD+0.29A^2+0.19B^2+0.17C^2+0.12D^2$ . (7)

315 In Equation (6), the constant term was unrelated to the factors, while the quadratic  
316 terms of  $A^2$ ,  $B^2$ ,  $C^2$ , and  $D^2$  positively affected the color difference, and consequently,  
317 the color difference value increased as these terms increased. The primary terms A, B,  
318 C, and D, as well as the interaction terms AB, AC, AD, BC, BD, and CD, did not have  
319 a significant effect on the color difference, therefore when these terms increased, the  
320 color difference decreased. Similarly, Equation (7) was consistent with the above  
321 analysis. The constructed model's dependability and importance were confirmed by the  
322 F-values (16.27 and 13.93). From the F-values, the effects of each factor on the color  
323 difference and gloss were established as follows: temperature (A) > enzymatic  
324 hydrolysis time (D) > enzyme concentration (C) > pH (B). The interaction between the  
325 independent variables can be understood by observing the P-value. From Table 4, the  
326 P-values (<0.0001) verified that the regression equations established with color  
327 difference and gloss as response values were significant. The linear terms' impacts on  
328 color difference and gloss were extremely significant ( $P < 0.01$ ), whereas interaction  
329 terms' effects on color difference and gloss were not substantial enough ( $P > 0.05$ ) to  
330 alter them. The quadratic terms ( $A^2$ ,  $B^2$ , and  $C^2$ ) exhibited an extremely significant ( $P$   
331  $< 0.01$ ) influence on the color difference and gloss. However, the effects of  $D^2$  on the  
332 color difference and gloss were respectively highly and generally significant. In  
333 addition,  $R^2$  (0.9421 and 0.9330) and adj- $R^2$  (0.8842 and 0.8660) values indicated that



334 the model fitted the experimental data well and minor changes (less than 0.1) cannot be  
 335 explained by the models. The lack of fitted values for the response surface models  
 336 (0.0558 and 0.3288) were not significant, indicating that the models were meaningful  
 337 for the optimization of the cleaning conditions. Therefore, the models could be used to  
 338 analyze and predict the effect of alkaline protease to remove paint.

339 Table 4 ANOVA of response surface model for the color difference and gloss.

Source	Color difference			Gloss		
	Mean square	F Value	Prob > F	Mean square	F Value	Prob > F
Model	54.71	16.27	<0.0001**	2.02	13.93	<0.0001**
A	11.25	46.85	<0.0001**	0.48	46.45	<0.0001**
B	5.08	21.17	0.0004**	0.12	11.61	0.0042**
C	6.95	28.93	<0.0001**	0.21	20.65	0.0005**
D	10.49	43.68	<0.0001**	0.40	39.03	<0.0001**
AB	0.11	0.45	0.5117	0.010	0.97	0.3419
AC	0.093	0.39	0.5437	$2.5 \times 10^{-3}$	0.24	0.6304
AD	0.47	1.95	0.1839	0.022	2.18	0.1622
BC	0.21	0.88	0.3638	$2.5 \times 10^{-3}$	0.24	0.6304
BD	0.18	0.75	0.4004	$2.5 \times 10^{-3}$	0.24	0.6304
CD	0.073	0.30	0.5903	0.010	0.97	0.3419
A <sup>2</sup>	15.20	63.28	<0.0001**	0.56	54.01	<0.0001**
B <sup>2</sup>	5.28	21.97	0.0003**	0.24	23.46	0.0003**
C <sup>2</sup>	5.25	21.85	0.0004**	0.18	17.79	0.0009**
D <sup>2</sup>	2.77	11.52	0.0044**	0.091	8.79	0.0102*
Residual	3.36			0.14		

Lack of fit	3.14	5.59	0.0558	0.12	1.67	0.3288
Pure error	0.22			0.028		
Cor total	58.07			2.16		

340 Notes: \*\* highly significant ( $P < 0.01$ ); \* significant ( $P < 0.05$ ).

#### 341 4.4.2 Analysis of surface responses

342 In order to optimize the key factors for optimal cleaning conditions, response  
343 surface (3D) and contour plots (2D) of the interaction between the four factors were  
344 produced in Figs. 7 and 8.

##### 345 4.4.2.1 Effect of independent variables on color difference

346 Six pairs of parameters influenced the color difference in Fig. 8. It can be observed  
347 in the 3D plots that when one variable was fixed, the color difference decreased and  
348 tended to stabilize as the other variable increased. The interactions of temperature with  
349 pH and enzyme concentration had no significant effect on the color difference due to  
350 the circularity of the contours in Figs. 7a and b. Given the elliptical shape of the  
351 contours in Figs. 7c and f, interactions concerning enzymatic hydrolysis time  
352 (temperature, pH, and enzyme concentration) and the interaction of pH with enzyme  
353 concentration on the color difference were significant. However, 3D surfaces of pH (Fig.  
354 8a), enzyme concentration (Fig. 8b) and enzyme hydrolysis time (Fig. 8c) were more  
355 flat than temperature, indicating that the effect of temperature on the color difference  
356 was the most obvious. The 3D surface of enzymatic hydrolysis time was steeper than  
357 pH (Fig. 8e) and enzyme concentration (Fig. 8f), while the 3D surface of pH was flatter  
358 than that of enzyme concentration (Fig. 8d), which indicated that the magnitude of the  
359 effect of three factors on gloss is enzymatic hydrolysis time > enzyme concentration >  
360 pH.

##### 361 4.4.2.2 Effect of independent variables on gloss

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362 Fig. 9 shows 3D and 2D plots for gloss as a function of the four independent factors.  
363 In the 3D plots, it can be noticed that for one variable to be stable, the gloss decreased  
364 and smoothed out with the increase of the other variable. By the tight circularity of the  
365 contours in Figs. 9a, b, and e, the interactions of temperature with pH and enzyme  
366 concentration, as well as the interaction between pH and enzymatic hydrolysis time,  
367 were irrelevant. From the elliptical contours in Figs. 9d, f, and c, the interactive effects  
368 of enzyme concentration with pH and enzymatic hydrolysis time, as well as temperature  
369 and enzymatic hydrolysis time, had a significant effect on gloss. The 3D surfaces of pH  
370 (Fig. 9a), enzyme concentration (Fig. 9b), and enzymatic hydrolysis time (Fig. 9c) were  
371 more flat than temperature, hence temperature had a more substantial impact on gloss  
372 than the other three factors. The 3D surface of enzyme concentration, both steeper than  
373 pH and flatter than enzymatic hydrolysis time, revealed that the order of their effects  
374 on glossiness was enzymatic hydrolysis time > enzyme concentration > pH.

#### 375 **4.4.3 Determination and experimental validation of optimal cleaning conditions**

376 The Design-Expert software's numerical optimization results revealed the most  
377 effective cleaning conditions: temperature: 42°C; pH of 9.5; enzyme concentration:  
378 1.92 mg/mL; and enzymatic hydrolysis time: 5.4 min. Three parallel experiments were  
379 conducted under optimal cleaning conditions to validate the developed model's  
380 dependability and accuracy. The results showed no significant differences ( $P > 0.05$ )  
381 between the color difference ( $2.25 \pm 0.26$ ) and gloss ( $0.1 \pm 0.1$  GU) obtained by parallel  
382 tests and the color difference (2.27) and gloss (0.2 GU) predicted by the models. As a  
383 result, the model was ideal and it can be used to remove surface paint using alkaline  
384 protease.

#### 385 **4.5 Validation experiments and analysis of surface roughness**

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386 According to the roughness measurements, the results indicated that the surface  
387 roughness values of blank stones and painted stones cleaned under optimal conditions  
388 were  $2.61 \pm 0.13 \mu\text{m}$  ( $R_{a1}$ ) and  $2.53 \pm 0.13 \mu\text{m}$  ( $R_{a2}$ ), respectively. The difference  
389 between the experimental results and the blank value was 8% (less than 10%), which  
390 indicated that the optimal cleaning conditions obtained from the model comply with the  
391 requirements for cleaning<sup>[56]</sup>.

#### 392 **4.6 Practical cleaning test of stone carvings in the Potala Palace**

393 The surface paint from stone relics H10-E09 of the Potala Palace was cleaned with  
394 alkaline protease under the optimum cleaning conditions. The unclean stone carvings  
395 were covered with white paint to conceal the information in Fig. 10a. It was found that  
396 after cleaning by alkaline protease that most of the paint was removed, indicating that  
397 alkaline protease had a good cleaning effect (Fig. 10b ). The results were in good  
398 agreement with those of the simulated cleaning test.

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399 **5. Conclusion**

400 We successfully selected the best cleaning agent from nine available biologically  
401 active options. The results show that alkaline protease has a good cleaning effect in  
402 terms of the removal of whitewash containing protein as a binder. This study provided  
403 a theoretical basis for the enzymatic cleaning of stone carvings and plays an essential  
404 role in protecting and repairing the Potala Palace stone carvings.

405

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431 **References**

432 [1] Harris C. The Potala Palace: Remembering to Forget in Contemporary Tibet, J.

433 South Asian Studies, 29 (2013), 61-75,

434 <http://doi.org/0.1080/02666030.2013.772816>.

435 [2] Dai R-C, Nabil W N N, Xu H-X. The history of saffron in China: From its origin

436 to applications, J. Chinese Medicine and Culture, 4 (2021), 228,

437 [http://doi.org/10.4103/CMAC.CMAC\\_38\\_21](http://doi.org/10.4103/CMAC.CMAC_38_21).

438 [3] Zuixiong L, Linyi Z, Li L, et al. Research on the modification of two traditional

439 building materials in ancient China, J. Heritage Science, 1 (2013), 1-11,

440 <http://doi.org/10.1186/2050-7445-1-27>.

441 [4] Fang S Q, Zhang H, Zhang B J, et al. The identification of organic additives in

442 traditional lime mortar, J. Journal of Cultural Heritage, 15 (2014), 144-150,

443 <http://doi.org/10.1016/j.culher.2013.04.001>.

444 [5] Gleize P, Müller A, Roman H. Microstructural investigation of a silica fume–

445 cement–lime mortar, J. Cement and Concrete composites, 25 (2003), 171-175,

446 [http://doi.org/10.1016/s0958-9465\(02\)00006-9](http://doi.org/10.1016/s0958-9465(02)00006-9)

- 
- 447 [6] Liu R Z, Zhang B J, Zhang H, et al. Deterioration of Yungang Grottoes: diagnosis  
448 and research, *J. Journal of Cultural Heritage*, 12 (2011), 494-499,  
449 <http://doi.org/10.1016/j.culher.2011.03.008>.
- 450 [7] Ke Z-B, Yang X-G, Lin Y, et al. Research on the Weathering and Influence of  
451 Prior Protective Coating above the Outdoor Stone Sculpture in Chongqing, *J.*  
452 *Spectroscopy and Spectral Analysis*, 37 (2017), 3229-3234,  
453 [http://doi.org/10.3964/j.issn.1000-0593\(2017\)10-3229-06](http://doi.org/10.3964/j.issn.1000-0593(2017)10-3229-06).
- 454 [8] Pintus A, Aragoni M C, Carcangiu G, et al. Density functional theory modelling of  
455 protective agents for carbonate stones: a case study of oxalate and oxamate  
456 inorganic salts, *J. New Journal of Chemistry*, 42 (2018), 11593-11600,  
457 <http://doi.org/10.1039/C8NJ01714J>.
- 458 [9] Zhang Y, Wu F S, Su M, et al. Spatial and temporal distributions of microbial  
459 diversity under natural conditions on the sandstone stelae of the Beishiku Temple  
460 in China, *J. International Biodeterioration & Biodegradation*, 163 (2021),  
461 <http://doi.org/10.1016/j.ibiod.2021.105279>.
- 462 [10] Li T X, Hu Y L, Zhang B J. Evaluation of efficiency of six biocides against  
463 microorganisms commonly found on Feilaifeng Limestone, China, *J. Journal of*  
464 *Cultural Heritage*, 43 (2020), 45-50, <http://doi.org/10.1016/j.culher.2019.11.006>.
- 465 [11] Centauro I, Cantisani E, Grandin C, et al. The Influence of Natural Organic  
466 Materials on the Properties of Traditional Lime-Based Mortars, *J. International*  
467 *Journal of Architectural Heritage*, 11 (2017), 670-684,  
468 <http://doi.org/10.1080/15583058.2017.1287978>.
- 469 [12] Villa P, Pollarolo L, Degano I, et al. A milk and ochre paint mixture used 49,000  
470 years ago at Sibudu, South Africa, *J. PloS one*, 10 (2015), e0131273,  
471 <http://doi.org/10.1371/journal.pone.0131273>.

- 
- 472 [13] Ventolà L, Vendrell M, Giraldez P, et al. Traditional organic additives improve  
473 lime mortars: New old materials for restoration and building natural stone  
474 fabrics, *J. Construction and Building Materials*, 25 (2011), 3313-3318,  
475 <http://doi.org/10.1016/j.conbuildmat.2011.03.020>.
- 476 [14] Kuckova S, Rambouskova G, Junkova P, et al. Analysis of protein additives  
477 degradation in aged mortars using mass spectrometry and principal component  
478 analysis, *J. Construction and Building Materials*, 288 (2021),  
479 <http://doi.org/10.1016/j.conbuildmat.2021.123124>.
- 480 [15] Hu W, Zhang K, Zhang H, et al. Analysis of polychromy binder on Qin  
481 Shihuang's Terracotta Warriors by immunofluorescence microscopy, *J. Journal of*  
482 *Cultural Heritage*, 16 (2015), 244-248,  
483 <http://doi.org/10.1016/j.culher.2014.05.003>.
- 484 [16] Rao H, Li B, Yang Y, et al. Proteomic identification of organic additives in the  
485 mortars of ancient Chinese wooden buildings, *J. Analytical Methods*, 7 (2015),  
486 143-149, <http://doi.org/10.1039/C4AY01766H>.
- 487 [17] Alonso E, Martinez-Gomez L, Martinez W, et al. Preparation and  
488 characterisation of ancient-like masonry mortars, *J. Advanced Composites*  
489 *Letters*, 11 (2002), 096369350201100105,  
490 <http://doi.org/10.1177/096369350201100105>.
- 491 [18] Pozo-Antonio J S, Rivas T, López A, et al. Effectiveness of granite cleaning  
492 procedures in cultural heritage: A review, *J. Science of The Total Environment*,  
493 571 (2016), 1017-1028, <http://doi.org/10.1016/j.scitotenv.2016.07.090>.
- 494 [19] Vergès-Belmin V, Rolland O, Jourd'heuil I, et al. Nd: YAG long Q-switched  
495 versus short free-running laser cleaning trials at Chartres cathedral, France, *J.*



- 
- 496 Studies in Conservation, 60 (2015), S12-S18,  
497 <http://doi.org/10.1179/0039363015Z.000000000202>.
- 498 [20] Chloros J, Salmon H, Talland V. Laser cleaning at the Isabella Stewart Gardner  
499 Museum, Boston, USA: Sixteen Roman sculptures, fourteen months, and three  
500 conservators, J. Studies in Conservation, 60 (2015), S41-S48,  
501 <http://doi.org/10.1179/0039363015Z.000000000206>.
- 502 [21] Carvalhão M, Dionísio A. Evaluation of mechanical soft-abrasive blasting and  
503 chemical cleaning methods on alkyd-paint graffiti made on calcareous stones, J.  
504 Journal of Cultural Heritage, 16 (2015), 579-590,  
505 <http://doi.org/10.1016/j.culher.2014.10.004>.
- 506 [22] Campos M I, Fortes S G, Pérez J L P. Influence of projection angle in  
507 sandblasting cleaning on detrietic stone materials in Architectural Heritage, J.  
508 Materiales de construcción, (2014), 10, <http://doi.org/10.3989/mc.2014.02113>.
- 509 [23] Moropoulou A, Tsiourva T, Bisbikou K, et al. Evaluation of cleaning procedures  
510 on the facades of the Bank of Greece historical building in the center of Athens,  
511 J. Building and environment, 37 (2002), 753-760, [http://doi.org/10.1016/S0360-  
512 1323\(01\)00058-0](http://doi.org/10.1016/S0360-1323(01)00058-0).
- 513 [24] Warscheid T, Braams J. Biodeterioration of stone: a review, J. International  
514 Biodeterioration & Biodegradation, 46 (2000), 343-368,  
515 [http://doi.org/10.1016/S0964-8305\(00\)00109-8](http://doi.org/10.1016/S0964-8305(00)00109-8).
- 516 [25] Young M E, Urquhart D, Laing R. Maintenance and repair issues for stone  
517 cleaned sandstone and granite building façades, J. Building and Environment, 38  
518 (2003), 1125-1131, [http://doi.org/10.1016/S0360-1323\(03\)00084-2](http://doi.org/10.1016/S0360-1323(03)00084-2).
- 519 [26] Hu W-W, Ge X-H, Ye W-R. Summarization of Airport Runway Lights Cleaning  
520 Methods, C. Journal, (2017), 114-117, <http://doi.org/10.2991/icsd-17.2017.17>.

- 
- 521 [27] Macchia A, Sammartino M P, Tabasso M L. A new method to remove copper  
522 corrosion stains from stone surfaces, *J. Journal of Archaeological Science*, 38  
523 (2011), 1300-1307, <http://doi.org/10.1016/j.jas.2011.01.005>.
- 524 [28] Delgado Rodrigues J, Valero J. A brief note on the elimination of dark stains of  
525 biological origin, *J. Studies in Conservation*, 48 (2003), 17-22,  
526 <http://doi.org/10.1179/sic.2003.48.1.17>.
- 527 [29] Han H, Zha J, Wang F, et al. Polyvinylamine Gel as a Cleaning Agent for  
528 Removing Mineral Crusts from Archaeologically Important Stone Artifacts, *J.*  
529 *Studies in Conservation*, (2021), 1-11,  
530 <http://doi.org/10.1080/00393630.2021.1935119>.
- 531 [30] Gervais C, Grissom C A, Little N, et al. Cleaning marble with ammonium citrate,  
532 *J. Studies in conservation*, 55 (2010), 164-176,  
533 <http://doi.org/10.1179/sic.2010.55.3.164>.
- 534 [31] Cremonesi P. Surface cleaning? Yes, freshly grated Agar gel, please, *J. Studies in*  
535 *conservation*, 61 (2016), 362-367,  
536 <http://doi.org/10.1179/2047058415Y.0000000026>.
- 537 [32] Porter J H, Pasian C, De Angelis R, et al. Cleaning of Oil-on-Stone Wall  
538 Paintings: Lessons Learned From Easel Painting Conservation, *J. Studies in*  
539 *Conservation*, 65 (2020), P254-P257,  
540 <http://doi.org/10.1080/00393630.2020.1753354>.
- 541 [33] De Witte E, Dupas M. Cleaning poultices based on EDTA, *C. Journal*, 3 (Year),  
542 1023-1031,
- 543 [34] Grassi S, Favaro M, Tomasin P, et al. Nanocontainer aqueous systems for  
544 removing polymeric materials from marble surfaces: A new and promising tool

- 
- 545 in cultural heritage conservation, *J. Journal of Cultural Heritage*, 10 (2009), 347-  
546 355, <http://doi.org/10.1016/j.culher.2008.10.003>.
- 547 [35] Valentini F, Diamanti A, Palleschi G. New bio-cleaning strategies on porous  
548 building materials affected by biodeterioration event, *J. Applied Surface Science*,  
549 256 (2010), 6550-6563, <http://doi.org/10.1016/j.apsusc.2010.04.046>.
- 550 [36] Mazzoni M, Alisi C, Tasso F, et al. Laponite micro-packs for the selective  
551 cleaning of multiple coherent deposits on wall paintings: The case study of  
552 Casina Famese on the Palatine Hill (Rome-Italy), *J. International  
553 Biodeterioration & Biodegradation*, 94 (2014), 1-11,  
554 <http://doi.org/10.1016/j.ibiod.2014.06.004>.
- 555 [37] Ranalli G, Zanardini E. Biocleaning on Cultural Heritage: New frontiers of  
556 microbial biotechnologies, *J. Journal of Applied Microbiology*, 131 (2021), 583-  
557 603, <http://doi.org/10.1111/jam.14993>.
- 558 [38] Ortega-Morales B O, Gaylarde C C. Bioconservation of historic stone  
559 buildings—An updated review, *J. Applied Sciences*, 11 (2021), 5695,  
560 <http://doi.org/10.3390/app11125695>.
- 561 [39] Abdelaal S, Sandu I C A. Assessment of protease in cleaning of bat blood patches  
562 from ancient Egyptian wall paintings and surface inscriptions, *J. International  
563 Journal of Conservation Science*, 10 (2019), 459-474,
- 564 [40] Segal J, Cooper D. THE USE OF ENZYMES TO RELEASE ADHESIVES, *C.  
565 Journal*, 2(1) (1977), 47-50, <http://doi.org/10.1080/03094227.1977.9638498>
- 566 [41] Han Y, Liu Z, Huang X, et al. The application of ELISA to the analysis and  
567 research of cementing materials in calligraphy of porcelain relics in Song  
568 Dynasty, *J. Microchemical Journal*, 159 (2020), 105530,  
569 <http://doi.org/10.1016/j.microc.2020.105530>.

- 
- 570 [42] Krizova I, Schultz J, Nemec I, et al. Comparison of analytical tools appropriate  
571 for identification of proteinaceous additives in historical mortars, *J. Analytical*  
572 *and bioanalytical chemistry*, 410 (2018), 189-200,  
573 <http://doi.org/10.1007/s00216-017-0709-8>.
- 574 [43] Huang H, Xu G, Liu X. Study on the Purity of Gold Leaf in a SO<sub>2</sub> Atmosphere at  
575 Ambient Temperature, *J. Materials*, 14 (2021), 2425,  
576 <http://doi.org/10.3390/ma14092425>.
- 577 [44] Benyounis K, Olabi A, Hashmi M. Effect of laser welding parameters on the heat  
578 input and weld-bead profile, *J. Journal of materials processing technology*, 164  
579 (2005), 978-985, <http://doi.org/10.1016/j.jmatprotec.2005.02.060>.
- 580 [45] Shu H, Song Y, Liu Q, et al. The study of rod-shaped TiO<sub>2</sub> composite material in  
581 the protection of stone cultural relics, *J. Green Processing and Synthesis*, 9  
582 (2020), 359-365, <http://doi.org/10.1515/gps-2020-0034>.
- 583 [46] Silva M M, Sanjad T a B C, Costa M L D, et al. Lime-based restoration paints:  
584 characterization and evaluation of formulations using a native species from the  
585 Amazon flora and PVA-based glue as additives, *J. Ambiente Construído*, 17  
586 (2017), 7-23, <http://doi.org/10.1590/s1678-86212017000300159>.
- 587 [47] Wang Y C, Shao M S, Zhang J K, et al. Quantitative evaluation of alteration and  
588 exfoliation in Jurassic sandstone, Chongqing Danzishi rock carvings, China, *J.*  
589 *Engineering Geology*, 292 (2021), <http://doi.org/10.1016/j.enggeo.2021.106277>.
- 590 [48] Klemm A J, Sanjeevan P, Klemm P. The effects of laser cleaning process on  
591 geometrical microstructure of cementitious composites, *J. Brittle Matrix*  
592 *Composites*, (2009), 323-334, <http://doi.org/10.1533/9781845697754.323>.
- 593 [49] Rabiller-Baudry M, Le Maux M, Chaufer B, et al. Characterisation of cleaned  
594 and fouled membrane by ATR—FTIR and EDX analysis coupled with SEM:

- 
- 595 application to UF of skimmed milk with a PES membrane, *J. Desalination*, 146  
596 (2002), 123-128, [http://doi.org/10.1016/S0011-9164\(02\)00503-9](http://doi.org/10.1016/S0011-9164(02)00503-9).
- 597 [50] Liu L, Wang Y, Peng C, et al. Optimization of the preparation of fish protein anti-  
598 obesity hydrolysates using response surface methodology, *J. International*  
599 *Journal of Molecular Sciences*, 14 (2013), 3124-3139,  
600 <http://doi.org/10.3390/ijms14023124>.
- 601 [51] Ramkumar A, Sivakumar N, Gujarathi A M, et al. Production of thermotolerant,  
602 detergent stable alkaline protease using the gut waste of *Sardinella longiceps* as a  
603 substrate: Optimization and characterization, *J. Scientific reports*, 8 (2018), 1-  
604 15, <http://doi.org/10.1038/s41598-018-30155-9>.
- 605 [52] Liu Y, Guo R. pH-dependent structures and properties of casein micelles, *J.*  
606 *Biophysical chemistry*, 136 (2008), 67-73,  
607 <http://doi.org/10.1016/j.bpc.2008.03.012>.
- 608 [53] Hagiwara H, Miyazaki K, Matuo Y, et al. Purification and characterization of  
609 alkaline protease and neutral protease from chromatin of rats, *J. Biochimica et*  
610 *Biophysica Acta (BBA)-Enzymology*, 660 (1981), 73-82,  
611 [http://doi.org/10.1016/0005-2744\(81\)90110-8](http://doi.org/10.1016/0005-2744(81)90110-8).
- 612 [54] Ma A, Ooraikul B. Optimization of enzymatic hydrolysis of canola meal with  
613 response surface methodology, *J. Journal of Food Processing and Preservation*,  
614 10 (1986), 99-113, <http://doi.org/10.1111/j.1745-4549.1986.tb00010.x>.
- 615 [55] Guan X, Yao H, Chen Z, et al. Some functional properties of oat bran protein  
616 concentrate modified by trypsin, *J. Food Chemistry*, 101 (2007), 163-170,  
617 <http://doi.org/10.1016/j.foodchem.2006.01.011>.

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618 [56] Grissom C A, Charola A E, Wachowiak M J. Measuring surface roughness on  
619 stone: back to basics, *J. Studies in Conservation*, 45 (2000), 73-84,  
620 <http://doi.org/10.1179/sic.2000.45.2.73>.

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## 627 **FIGURE LEGENDS**

628 **Fig. 1** Sampling location map of Potala Palace: the bottom left side of H10-E11 stone  
629 carving (①), right side of stone carving E37-E38 (②), and right side of H10-E39  
630 stone carving (③) were sampling points, respectively.

631 **Fig. 2** The field images of the sampling points in H10-E39. (a: outer layer, b: middle  
632 layer, c: bottom layer).

633 **Fig. 3** Images of the powdered paint samples used in the experiments.

634 **Fig. 4** Standard curve of casein.

635 **Fig. 5** Corrosion of simulated stones by different cleaning agents. Different lowercase  
636 letters after the data in the same column indicate that one-way ANOVA of the  
637 mean values differs significantly ( $P < 0.05$ ).

638 **Fig. 6** The color difference of simulated stones after being cleaned by different  
639 cleaning agents. Different lowercase letters after the data in the same column  
640 indicate that one-way ANOVA of the mean values differs significantly ( $P <$   
641  $0.05$ ).

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642 **Fig. 7** Influence of various factors on color difference and luster of simulated stone.

643 Different lowercase letters after the data in the same column indicate that one-

644 way ANOVA of the mean values differs significantly ( $P < 0.05$ ).

645 **Fig. 8** Response surface plots (3D and 2D) for the effect of temperature, pH, enzyme

646 concentration, and enzymatic hydrolysis time on the color difference.

647 **Fig. 9** Response surface plots (3D and 2D) for the effect of temperature, pH, enzyme

648 concentration, and enzymatic hydrolysis time on gloss.

649 **Fig. 10** The Potala Palace stone carvings practical cleaning application.